

TIMELINE

Self-organization in cell biology: a brief history

Eric Karsenti

Abstract | Over the past two decades, molecular and cell biologists have made important progress in characterizing the components and compartments of the cell. New visualization methods have also revealed cellular dynamics. This has raised complex issues about the organization principles that underlie the emergence of coherent dynamical cell shapes and functions. Self-organization concepts that were first developed in chemistry and physics and then applied to various morphogenetic problems in biology over the past century are now beginning to be applied to the organization of the living cell.

One of the most fundamental problems in biology concerns the origin of forms and their associated functions. This has been a long-lasting question in developmental biology, but similar questions must also be addressed at the cellular level. Since the discovery of the structure of DNA, the genome has often been thought of as the overriding architect: a given combination of genes that determines the phenotype through a linear chain of causal events. The problem is that embryogenesis and dynamic cell forms and functions emerge from multiple molecular interactions and interconnected regulatory feedback loops^{1–4}. Moreover, many parameters, such as physical constraints and collective behaviours, are not under the direct control of the genome. Therefore, we cannot hope to explain cell morphogenesis, for example, by invoking simple linear chains of causal events that link genes to phenotypes^{5–7}.

It seems that the philosopher Kant was the first to define life as a “self-organized, self-reproducing” process (TIMELINE). Through pure reasoning, he defined life as the emergence of functions by self-organization. He said that in an organism, every part owes its existence and origin to that of the other parts, with the functions that are attributed to a complete living organ or organism emerging from the properties of the parts and of the whole. He defined

this complex state of living matter as a self-organized end^{8–10}. This led him to question the validity of using the causality principle of classical physics to explain life, and to suggest that a new kind of science would be required to study how purpose and means are intricately connected⁸.

The new science he was talking about did emerge much later, from observations and studies made by chemists and physicists who discovered new, more complex forms of causality than what Kant had foreseen^{11,12}. Ironically, although Kant attempted to characterize life as a self-organization process in opposition to non-living matter, the first well-defined concepts and observations of self-organized processes came from theoretical considerations by Lotka^{13,14}, from chemistry by Bray¹⁵ and from the Belousov–Zhabotinsky reaction^{16–19} (TIMELINE). Chemical oscillations emerged from reaction–diffusion processes that were formalized in mathematical terms by Kolmogorov *et al.*²⁰ in the 1930s and by Turing in the 1950s, who predicted that

“It was clear from the outset that the emergence of dynamical organization observed in physical and chemical systems should be of importance to biology...”

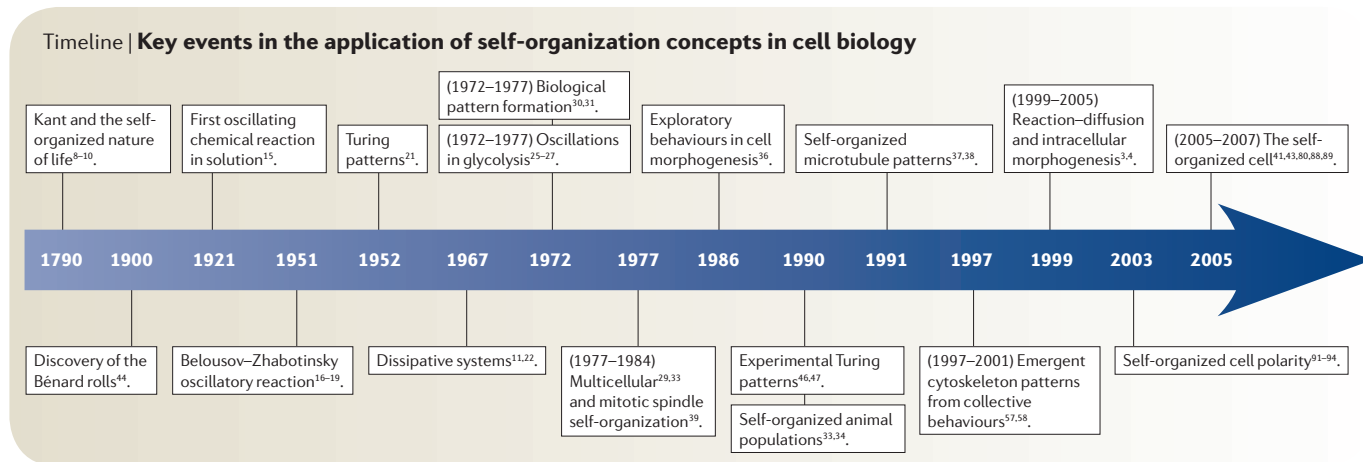
steady-state spatial patterns could also arise from such processes in living systems²¹. The full formalization of the nature of self-organization processes came from the work of Prigogine on instabilities and the emergence of organization in ‘dissipative systems’ in the 1960s^{22–24}, and from Haken who worked on similar issues under the name of synergetics¹¹ (TIMELINE).

It was clear from the outset that the emergence of dynamical organization observed in physical and chemical systems should be of importance to biology, and scientists who are interested in the periodic manifestations of life and developmental biology have been actively working in this field^{19,25–29}. From a more general point of view, Kauffman built on the ideas of Prigogine and Haken in an attempt to explain the origin of order in biology^{30–32}. Self-organization was also invoked to explain the formation of regular patterns in the fur of animals and the collective behaviour of organisms in ant colonies, termite nest building, schools of fish and flocks of birds^{33,34}. The importance of self-organization processes in molecular cell biology began to be recognized in the 1980s and 1990s^{1,35–40}, but only really started to gain momentum recently^{6,41–43} (TIMELINE).

In the following article, I do not deal with developmental biology issues but specifically focus on how self-organization principles and mechanisms (BOX 1) can help to understand subcellular and whole-cell morphogenesis. I first summarize the essence of the theory of self-organization in physico-chemical systems in simple terms. I then show how this concretely applies to some examples of cell organization and function.

Self-organization concepts

The initial definition of self-organization by Kant as a characteristic of living systems implied the existence of a loop between organization and function. A simpler definition used by modern scientists is that dynamic organization emerges from the collective behaviour of ‘agents’, the individual properties of which cannot account for the properties of the final dynamic pattern. This definition is more general and has the



advantage of being applicable to systems that do not necessarily acquire a function, even though they become dynamically organized. The understanding of the emergence of function can be studied separately. The other advantage of this definition is that it establishes how self-organized dynamical systems should be studied: the goal of the science of self-organization is to identify the principles and mechanisms by which an ensemble of agents in interaction evolves towards a particular dynamical temporal or spatial pattern.

Origins in thermodynamics. At first sight, the spontaneous emergence of order in the universe contradicts the second law of thermodynamics: a thermodynamically closed system settles in the most disordered state (that is, the state with the highest entropy, when molecules occupy all of the space randomly; FIG. 1a). Ordered states can and do emerge at thermodynamic equilibrium (for example, crystals, lipid bilayers, molecular complex formation), but they are static.

In a thermodynamically open system that receives energy from the outside, the

energy that flows through it can be used to decrease its entropy (that is, generate order). Molecules can suddenly organize themselves in dynamic patterns. Bénard rolls^{44,45} (longitudinal cylinders of liquid molecules that form precise and stable dynamic patterns) represent such an example and were called ‘dissipative structures’ by Prigogine²² (FIG. 1b).

Collective behaviour and the Bénard rolls.

Bénard rolls form when a liquid is heated from below, which generates a temperature gradient. Molecules at the bottom of the container are more agitated than at the top, creating a lighter layer of liquid than that at the top. Roll formation results from local instabilities that lead the system to break its symmetry when molecules start to behave collectively. This happens at different critical temperatures for different fluids, but always occurs when all of the parameters balance each other so that they satisfy a universal number, called the Rayleigh number, which equals 1708 under specific conditions³³.

The system can generate rolls that, at a given position in the container, move clockwise or counter-clockwise with alternate orientation. When the temperature is raised to the critical value, the system bifurcates between two alternative steady states (FIG. 1b). The bifurcation is based on local fluctuations that occur at the critical temperature and is almost irreversible. This comes from the fact that the molecules in the liquid begin to behave collectively, all moving up together on one side of the roll and downwards on the other (FIG. 1b). A long-range correlation has been established between the molecules of the system: the whole pattern of rolls in the container emerges from the collective properties of the molecules in the fluid and the geometry of the container, and cannot be predicted from the properties of any of its parts.

Box 1 | Self-organization concepts and mechanisms

Self-organization occurs when elements interact dynamically with each other to generate a system that acquires emergent properties that cannot be directly predicted from the individual properties of the elements. This only happens when the system dissipates energy.

Principles	Mechanisms	Examples in the cell
Thermodynamics: non-equilibrium thermodynamics.	Thermal, chemical or other energy dissipation that is associated with dynamic pattern formation.	ATP consumption coupled to dynamic pattern formation.
Symmetry breaking: occurs when a system switches from one symmetry level to another.	Gravity, temperature or chemical gradients, local fluctuations.	Intrinsic asymmetry of agents, nonlinear reactions, stereospecific localization of enzymes, pre-existing structures.
Emergence: a new property that arises from the collective behaviour of agents.	Collective effects and reactions that lead to systems properties.	Cytoskeleton behaviour, enzymatic oscillators, functional networks.
Robustness: the system evolves towards a steady state that constrains its agents to remain within this steady state.	Feedback loops, physical or collective constraints.	Reaction networks with two-state systems, such as kinase–phosphatase or small G-protein systems, combined with feedback loops. Physical, chemical and collective constraints.
Bifurcation: the system moves from one steady state to another when a specific parameter varies around a critical value.	Local instabilities at critical parameter values, nonlinearities.	Toggle switches between network states, switches between collective dynamics states.

Feedback loops and spatio-temporal patterns. In the example of Bénard rolls, order emerges from collective physical interactions. In Belousov–Zhabotinsky reactions and Turing patterns, order emerges from the nonlinear kinetic properties of chemical reactions: that is, from a combination of diffusion and feedback loops in the reaction system. Turing²¹ showed that a system of reactants that are initially homogeneously distributed in a solution can generate products that segregate in spatial patterns if certain conditions are met. The product of a reaction should act as a short-range positive activator of its own production while activating the production of an inhibitor that diffuses much faster^{46,47}. This forms a local positive feedback loop and a long-range negative feedback loop. The symmetry of the solution is broken by diffusion-driven instabilities that get amplified by the feedback loops. Waves or steady-state patterns can emerge^{48,49}. Again, the system can bifurcate between different states. In biology, such reaction–diffusion processes have obvious morphogenetic potential^{50,51}. The problem is to identify them and understand how biological molecules interact to form the appropriate reaction networks. Biologists have been trying to find reaction–diffusion mechanisms that fit the Turing model exactly, especially in developmental biology^{29,28}. Because biological molecules are complex, various types of reaction–diffusion processes that involve enzymatic reactions may occur — these might have powerful morphogenetic properties without having the exact characteristics of the Turing equations^{3,4}. In addition, combinations of positive and negative feedback loops or other properties, such as ultrasensitivity and cooperativity, provide powerful sources of nonlinearities that can lead to the building of enzymatic oscillators or steady-state patterns^{27,52}. Therefore, when trying to understand the self-organization properties of cells or embryos, it is more important to simply treat them as nonlinear dynamical systems than to try to find exactly what Turing had predicted.

Self-organization and the cell

A large part of cell organization depends on self-assembly processes that do not involve energy dissipation (thermodynamic equilibrium). But in the living cell, self-organization processes also occur through the dissipation of ATP or GTP. In fact, the dynamic order of the cell results from a combination of complex stereospecific interactions (deterministic self-assembly)

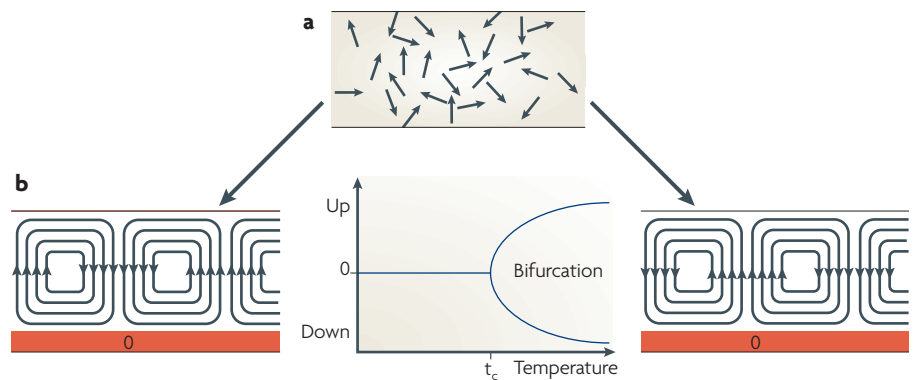


Figure 1 | **Bénard rolls.** **a** | In a liquid layer, molecules are agitated by thermal motion. **b** | The molecules in the liquid layer are heated from below (red zone) and self-organize into rolls (drawn in cross-section) when the temperature reaches a critical value (t_c). At this value, the molecules start to move collectively either up or down at point 0, which determines the alternative orientation of the rotation of the rolls throughout the layer. The orientation of the rotation choice is unpredictable and determined by local fluctuations at t_c .

and extremely varied dynamical interactions between molecules that require energy dissipation (self-organization). Here, I give some recent examples in which the problem of cellular self-organization has been addressed — from well-studied systems to more complex and less understood ones.

“...in the living cell, self-organization processes also occur through the dissipation of ATP or GTP.”

Patterns and oscillators from collective behaviours.

In 1990, Tabony and Job³⁷ published a paper in which they showed that pure microtubule solutions self-organized into stripes when they were incubated for several hours at 33°C, and noticed the analogy with patterns formed in the Belousov–Zhabotinsky reaction. Computer simulations suggest that the patterns may arise through a reaction–diffusion mechanism⁵³. However, different interpretations of these observations have been proposed and have suggested that, instead, the patterns arise through collective effects that result from dynamic instability coupled to microtubule buckling⁵⁴. In fact, the stripes are apparently made of aligned microtubules and, interestingly, the patterns formed are affected by boundary conditions⁵⁵. These properties may contribute to complex patterns that are formed *in vivo*⁵⁶, but microtubule pattern formation in cells often requires both the regulation of microtubule dynamics and the contribution of motors.

In 1991, Verde and colleagues showed that randomly nucleated microtubules in frog egg extracts organized themselves with the minus-end motor dynein to form steady-state asters³⁸. This was further developed *in vitro*⁵⁷ and then *in silico*⁵⁸. Computer simulation analyses showed that various patterns can emerge from interactions between similar components as a function of their physical properties, complexity and concentration^{58,59}. Asters, vortices, antiparallel microtubule bundles and spindle networks were observed (FIG. 2a). In these systems, symmetry breaking comes from the asymmetric shape of the parts: the tubulin molecule is asymmetric⁶⁰ and the motors move towards one microtubule end or the other⁶¹. Therefore, the system has a built-in symmetry-breaking mechanism (FIG. 2a). Self-organization comes from the collective behaviour of the motors and microtubules, with the energy being dissipated by the motors as they move along microtubules.

Recently, these observations have been extended to the acto-myosin system⁶². Just as for microtubules, myosin II crosslinks actin filaments and the system self-organizes into various patterns including rings. Physicists call these systems active gels (as opposed to gels that are made of polymers that do not contain motors) and have developed a theory that predicts the behaviour of these active gels⁶³. For an example of structures that are generated by self-assembly at thermodynamic equilibrium, see the study by Haviv *et al.*⁶⁴, who obtained self-assembled actin stars that formed in a passive gel of actin and actin regulators.

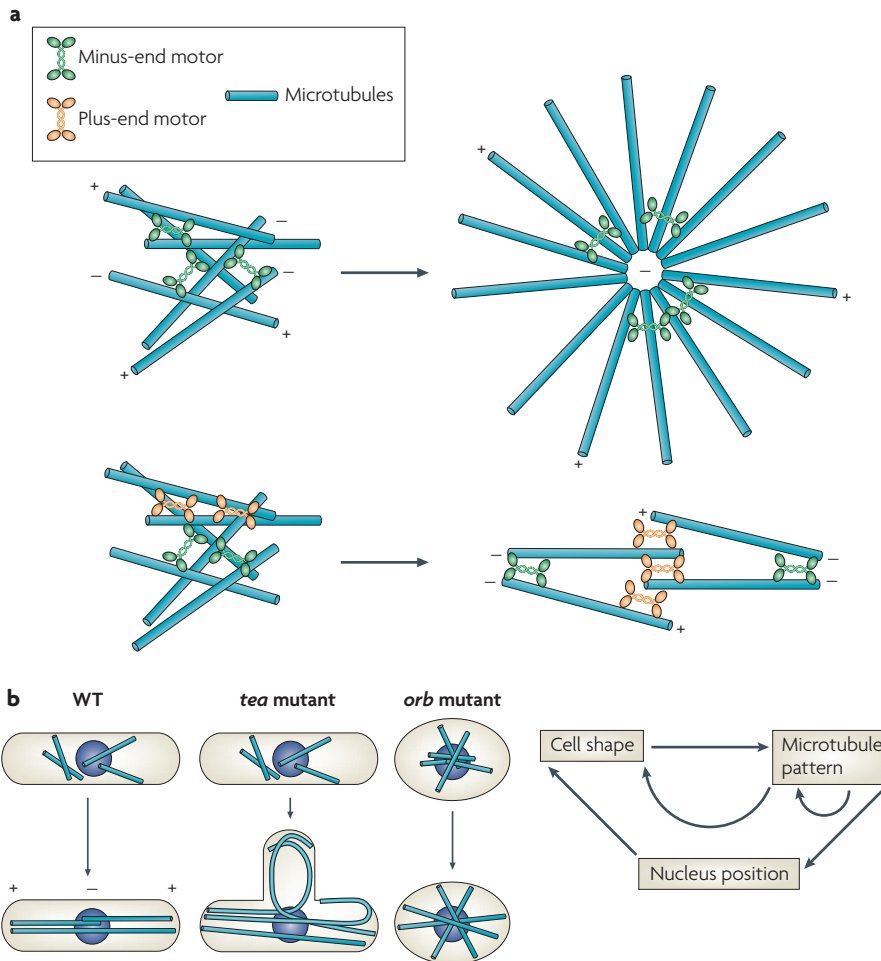


Figure 2 | Examples of self-organized microtubule patterns and cell shapes. **a** | Self-organization of mixtures of microtubules and motors. A minus-end motor can, under certain conditions, crosslink microtubules and focus the minus ends to form asters (top). A mixture of minus-end and plus-end motors can form various patterns. An antiparallel pattern with overlapping plus ends⁵⁹ is shown (bottom). **b** | Self-organization of microtubule patterns and cell shape in *Schizosaccharomyces pombe*. The elongated shape of *S. pombe* forces microtubules to align because microtubules depolymerize when they reach the tips of the cells. Motors (together with crosslinking molecules) force microtubules to form antiparallel bundles with the minus ends at the cell centre⁸⁷. In tip-elongation-aberrant (*tea*) mutants, microtubules keep growing when they reach the cell tips. Because tip-promoting factors move towards microtubule plus ends and because, in this case, microtubules curl along cell edges, additional growth tips can form, thereby generating T-shaped cells⁸⁴. Cells that are mutated in the Ser–Thr protein kinase *Orb6* (*orb* mutants) have a round shape and microtubules cannot organize into long bundles⁸³. This shows that self-organization of microtubules and the cell cortex feed back on each other to generate a self-organized dynamic cell shape. The circularity of this process is shown on the right of the figure.

An important aspect of self-organization in living systems concerns the formation of oscillators. They arise from the collective behaviour of cytoskeletal systems or from enzymatic networks (see below). Kruse and Jülicher have reviewed this field recently⁴², and the first theoretical treatment of the emergence of spontaneous oscillations of collective molecular motors was published in 1997 (REF. 65). The principle is relatively simple: a collection of motors can lose its grip on the filaments in a cooperative manner before

rebinding, which leads to dynamic instability of the force–velocity relationship and collective periodic binding and unbinding. This demonstrates how nonlinear collective effects can lead to periodic temporal patterns (for many other examples, see REF. 42).

Patterns and oscillators from enzymatic feedback loops. Substantial work on oscillators has already been mentioned above, but relatively little has been said about pattern formation inside cells. Here, I take the cell

cycle in eukaryotes as an example because it is probably the best-characterized system. (Further examples in biology in general and in cells more specifically can be found in REFS 27, 29, 42, 66.)

In eukaryotic cells, an oscillator coordinates DNA replication with chromosome segregation. The cyclin-dependent kinase (CDK) oscillator^{67,68} (FIG. 3a) self-organizes through the permanent synthesis of one protein, cyclin, which triggers intertwined positive and negative feedback loops^{68–74}. The CDK oscillator is a truly self-organized temporal pattern because it is entirely autonomous. The principle of its mechanism is analogous to a Belousov–Zhabotinsky reaction, except that it is built of much more complex molecules. It has two stable states — bifurcation between the two states is triggered when cyclin reaches a threshold concentration and when it is degraded. It works because there is a long time-delay built into the negative feedback loop that leads to cyclin degradation⁷².

The function of this oscillator is to change abruptly the cytoplasmic state of the cell. When *CDK1* is inactive, a nucleus assembles and DNA replicates, and when it is active, a mitotic spindle assembles. In both cytoplasmic states, chromatin breaks the spatial symmetry of the cytoplasm by the local accumulation of regulatory factors (through stereospecific targeting) that set off a series of (enzymatic) reaction–diffusion processes, which leads to the emergence of a nucleus in interphase and a spindle in metaphase^{3,75} (FIG. 3b). Therefore, this reaction–diffusion process contributes to the formation of spatial patterns inside the cytoplasm. The whole cell cycle in eukaryotes can be seen as being based on the principle of self-organization by reaction–diffusion, both temporally and spatially. I say reaction–diffusion, although it is important to realize that none of these processes are true Turing patterns. Indeed, the symmetry is not broken by spontaneous instabilities, but rather by deterministic effects (cyclin synthesis for the oscillator, and stereospecific targeting of a small G-protein exchange factor to chromatin for nuclear and spindle assembly).

The emergence of functions. Above, I have described purified cytoskeletal molecules that can self-organize into patterns as a result of their collective behaviour. In isolation, these patterns have no function because they have nothing to act on. In the cell, however, this might be different. During mitosis, for example, chromosomes induce the assembly of a spindle that acts on the chromosomes

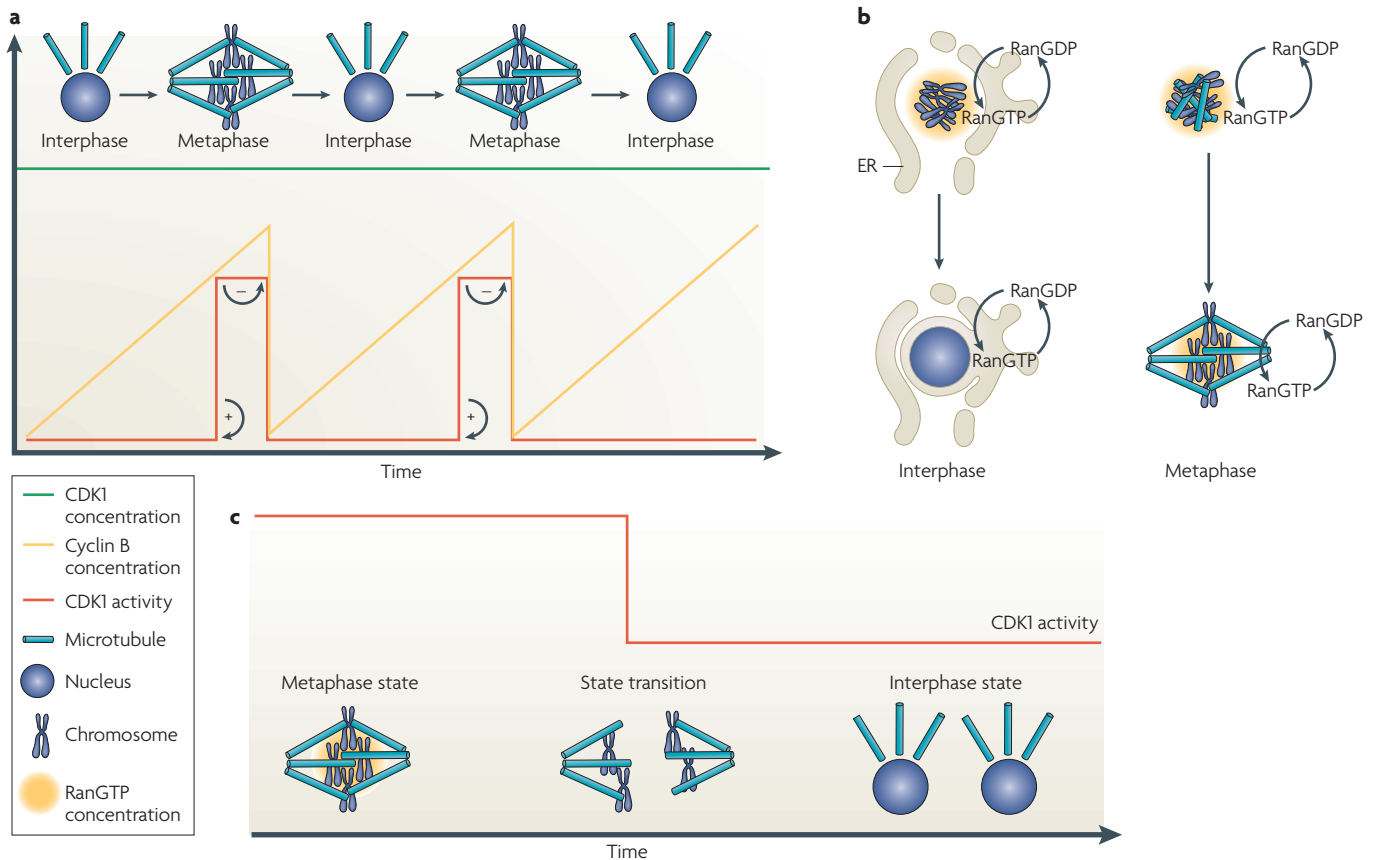


Figure 3 | Examples of self-organized cell-cycle processes. **a** | In eukaryotes, the timing of the cell cycle is determined by an oscillator that is driven by the accumulation of cyclin B, which binds to cyclin-dependent kinase-1 (CDK1). When the concentration of the cyclin B–CDK1 complex reaches a threshold, it triggers a positive feedback loop that leads to the abrupt activation of CDK1. When CDK1 activity reaches a threshold level, it triggers the delayed degradation of cyclin B (negative feedback), which results in the onset of anaphase. **b** | Chromatin generates a gradient of RanGTP that, through a series of complex reactions, triggers nuclear

assembly in interphase and spindle assembly in metaphase. The local production of RanGTP is determined by a reaction–diffusion mechanism. **c** | The transition between metaphase and interphase corresponds to a bifurcation between two steady states; that is, nuclear components are in two discrete dynamic interaction states that respond to the presence of DNA by self-organizing into either a nucleus or a spindle. When CDK1 is inactivated at the end of metaphase, the cytoplasm moves through a transient state. This is when chromosomes are segregated. ER, endoplasmic reticulum.

themselves. Therefore, the chromosomes trigger the self-organization of a pattern (the spindle) that acquires the function of segregating them⁷⁶. This is exactly the loop between organization and function that Kant was looking for and there is nothing mysterious about it — so, we can now understand the structure of this logic because we understand many of the underlying mechanisms.

There are three interlocked, self-organized systems that constitute the whole cell cycle: the oscillator (temporal self-organization), the spindle and the nucleus (spatial self-organization), and each subsystem has its own specific function. However, it is interesting to note that chromosome segregation does not occur while the spindle is at steady state, but during anaphase; that is, while the cell bifurcates back towards interphase, when a new steady state is established and a nucleus has reassembled (FIG. 3c).

The functional organization of the nucleus in interphase is another interesting example of self-organization in which function and structure are interdependent, as described by Misteli, Glick and Cook^{41,43,77–80}, although the exact mechanisms involved are still under investigation.

Let's take just two more examples of the emergence of function by self-organization. We have seen above that actin and myosin can form self-organized contractile rings *in vitro*⁶². Again, these rings have no function. However, *in vivo*, similar rings self-organize, triggered by the small G protein *RhoA* that is locally activated at the plasma membrane at the mid-zone of the dividing cell. This contractile ring functions to cut the cell in two⁸¹. In fact, this system works exactly like chromosome-induced spindle assembly, both logically and mechanistically: the final

function of the self-organized cytoskeletal structure is triggered by the cell domain on which it should act.

The second example concerns the spontaneous beating of axonemes that results from the self-organization of bending waves, which are generated by the collective effects of motor activity and bending elasticity of microtubules^{42,82}. Axoneme beating (the function) emerges from the self-organization properties of the system, which itself results from the interaction between dynein and microtubules.

Self-organized cells. The loops that link self-organization and function are also found when we start to look at the self-organization of large systems such as whole cells. In *Schizosaccharomyces pombe*, for example, the self-organization of microtubule bundles is required to maintain the

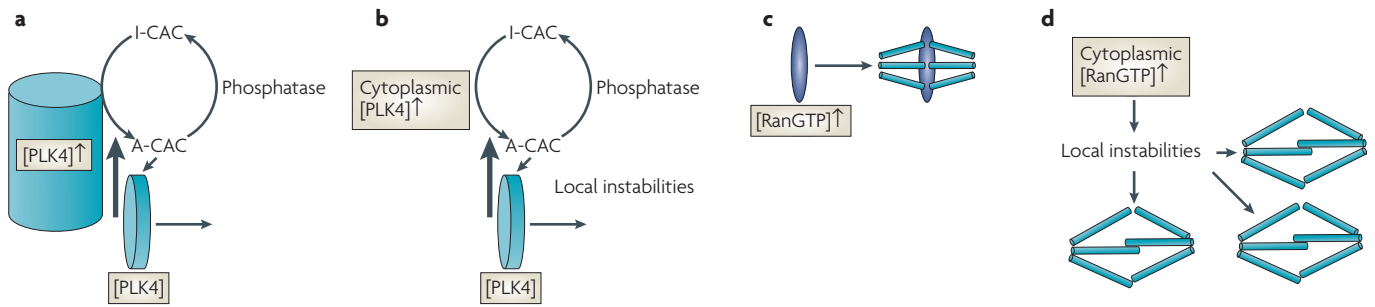


Figure 4 | Self-organized versus templated pattern transmission. **a** | A model of centriole duplication or spontaneous formation. The Polo-like kinase PLK4 is present in very low amounts in the cytoplasm but binds strongly to existing centrioles (turquoise) at the time of centriole duplication¹⁰². Because its substrate (centriole assembly complex; CAC) and the opposing phosphatase are in solution, inactive CAC (I-CAC) becomes phosphorylated and activated (A-CAC) just around centrioles, which leads to the assembly of new centrioles that bind more PLK4. This, in turn, leads to the autocatalytic assembly of a new centriole. **b** | In the absence of pre-existing

centrioles, PLK4 needs to be present in high concentrations in the cytoplasm to counterbalance the phosphatase enzyme of CAC. In this case, local instabilities can lead to localized assembly of A-CAC that could trigger the autocatalytic assembly of centrioles. This is based on the same idea as that of Turing: a local positive feedback loop coupled to long-range inhibition (the phosphatase). **c** | The previous scenario is analogous to the mechanism of RanGTP-induced local spindle assembly in the presence of a localized GTP exchange factor on chromatin, or **d** | global spontaneous spindle assembly if RanGTP is overexpressed throughout the cytoplasm.

simple cylindrical morphology. But the reverse is also true: cell shape is important to determine the morphology of microtubule bundles that self-organize through dynamic instability, the action of motors and associated molecules, and interaction with the cell periphery^{83–87}. Again, we find a loop between the whole and the parts^{88,89} (FIG. 2b).

In *S. pombe*, we really begin to understand how the nucleus, the microtubule system and cell shape form a circular integrated self-organized system in which none of the elements comes first (FIG. 2b). This is also a beautiful example of modular self-organization in which one versatile system (the microtubule system) interacts in various ways with other parts of the cell that are themselves self-organized, to form a whole. Chemotaxis and cell polarization are also interesting in this context. Cell polarization can occur in the absence of any cue through spontaneous symmetry breaking^{90,91}, involving instabilities in small G proteins, phosphorylation and cytoskeletal networks. This self-organized process can become functional for the cell by responding in a directional way to signalling gradients^{90–95}.

De novo versus templated. A discussion of self-organization brings us naturally to the old problem of templated versus *de novo* formation of organelles. It is clear that structural heredity exists: the structure and shape of membranes, the Golgi apparatus, centrioles and even whole cytoskeleton patterns pass from one generation to the next. But how are these structures transmitted? Does this occur through some sort of template that duplicates and on which new structures grow, or do these structures assemble

periodically *de novo* through instability-driven self-organization by virtue of the physical properties of their components?

A good example is the centriole. There have been many discussions about the templated duplication of centrioles because of their beautiful geometry and the mechanism of duplication at an exact angle to each other^{96–99}. However, centrioles do arise *de novo* in many cases¹⁰⁰, and the recent discovery of Polo-like kinase-4 (PLK4), which is essential for centriole duplication, indicates how this *de novo* formation may occur¹⁰¹. Indeed, overexpression of PLK4 triggers the formation of ectopic free centrioles, which suggests that under some conditions, pre-existing centrioles act as a concentration spot for PLK4 (that is, it binds to centrioles)¹⁰². If the opposing activity of a phosphatase is present throughout the cytoplasm, PLK4 may locally phosphorylate key substrates around pre-existing centrioles, leading to local self-organization of centriolar components through a reaction–diffusion mechanism (FIG. 4a). In the absence of pre-existing centrioles, an excess of PLK4 could phosphorylate the same substrate by opposing the phosphatase globally and, through local instabilities, trigger spatially restricted positive feedback loops that lead to local assembly of the nucleating agents that are required to initiate centriole self-organization (FIG. 4b). This would be similar in principle to chromatin-induced spindle

“...it now seems possible that the Golgi apparatus self-organizes from the ER in response to its very function.”

self-organization⁷⁶ (FIG. 4c,d). Indeed, in this latter case, when chromosomes are present they also induce the self-organization of spindles around them. However, on overexpression of the signalling molecules that are normally activated by chromosomes, spindles self-organize randomly in the cytoplasm in the absence of chromosomes¹⁰³ (FIG. 4d).

The case of the Golgi apparatus is also interesting. Again because of its unique structure, it has been proposed that the Golgi duplicates during cell division through a templated process¹⁰⁴. However, further work suggested that, in fact, new Golgi buds out from the endoplasmic reticulum (ER) at sites that are apparently specified by the basal body^{105,106}, and it now seems possible that the Golgi apparatus self-organizes from the ER in response to its very function^{41,107–109}.

The essence of life and evolution

There is a tendency for engineers who enter the field of biology to speak of design. Design does not exist in living matter (unless we believe in creationism). Nobody ‘thought’ through the advantage of positive and negative feedback loops to build the cell-cycle oscillator in *Xenopus laevis*. It just springs from a mixture of gene products that interact dynamically with each other, as in Belousov–Zhabotinsky reactions.

Something incredibly important for the understanding of the origin of life and evolution is emerging here: self-organization principles tell us that if there is an ensemble of products that can interact dynamically to reach a functional steady state, they will do so robustly at least under certain conditions³¹. Suddenly, life becomes much less improbable, as Kauffman suggested³⁰.

The principles that are associated with self-organization processes tend to indicate that the driving force behind the diversity of life and its evolution is not mainly selection. Instead, it may derive largely from the intrinsic properties of living matter and the combination of various self-organized functional modules. The maximum diversity of life is probably represented by the parameter space within which dynamic interactions between the whole and the parts robustly produce a cell or an organism that can survive in a given environment. In a sense we are moving back to the views of D'Arcy Thompson, who thought that biologists overemphasize the role of evolution over that of physical laws in the origin of growth and form⁵.

Conclusion

Studying self-organization processes in cell biology forces us to focus on principles and collective behaviours rather than on single molecules. This makes it necessary to use mathematics and physics as well as computer simulations to analyse the often counter-intuitive properties of dynamical systems. This also forces us to clarify the difference between principles and mechanisms (BOX 1).

Eric Karsenti is at the Cell Biology and Biophysics Unit,
European Molecular Biology Laboratory,
Meyerhofstrasse 1, 69117 Heidelberg, Germany.
e-mail: karsenti@embl.de
doi: 10.1038/nrm2357

- Babloyantz, A. *Molecules, Dynamics, and Life: An Introduction to Self-Organization of Matter* (Wiley, New York, 1986).
- Murray, J. Discussion: Turing's theory of morphogenesis — its influence on modelling biological patterns and form. *Bull. Math. Biol.* **52**, 119–152 (1990).
- Bastiaens, P., Caudron, M., Niethammer, P. & Karsenti, E. Gradients in the self-organization of the mitotic spindle. *Trends Cell Biol.* **16**, 125–134 (2006).
- Kholodenko, B. N. Cell-signalling dynamics in time and space. *Nature Rev. Mol. Cell Biol.* **7**, 165–176 (2006).
- Thompson, D. W. *On Growth and Form* (Cambridge Univ. Press, 1942).
- Kirschner, M., Gerhart, J. & Mitchison, T. Molecular "vitalism". *Cell* **100**, 79–88 (2000).
- Kurakin, A. Self-organization versus watchmaker: ambiguity of molecular recognition and design charts of cellular circuitry. *J. Mol. Recognit.* **20**, 205–214 (2007).
- Kant, E. *Critique de la Faculté de Juger* (Gallimard, Paris, 1985) (in French).
- Van de Vijver, G. *Self-Organization and Emergence in Life Sciences* (Springer, Dordrecht, 2006).
- Fox Keller, E. Contenders for life at the dawn of the twenty-first century: approaches from physics, biology and engineering. *Interdiscip. Sci. Rev.* **32**, 113–122 (2007).
- Haken, H. Nonequilibrium phase transitions and self-organisation in physics, chemistry, and biology. In *Synergetics: An Introduction* (Springer, Berlin, 1977).
- Prigogine, I. & Stengers, I. *Order Out of Chaos* (Bantam, Toronto, 1984).
- Lotka, A. J. Contributions to the theory of periodic reactions. *J. Phys. Chem.* **14**, 271–274 (1910).
- Lotka, A. *Elements of Physical Biology* (Williams and Wilkins, Baltimore, 1925).
- Bray, W. A periodic reaction in homogeneous solution and its relation to catalysis. *J. Am. Chem. Soc.* **43**, 1262–1267 (1921).
- Belousov, B. [A periodic reaction and its mechanism]. *Compilation of Abstracts on Radiation Medicine* **147**, 145 (1959) (in Russian).
- Zhabotinsky, A. [Periodic processes of malonic acid oxidation in a liquid phase.]. *Biofizika* **9**, 306–311 (1964) (in Russian).
- Zhabotinsky, A. M. & Zaikin, A. N. Autowave processes in a distributed chemical system. *J. Theor. Biol.* **40**, 45–61 (1973).
- Tabony, J. Historical and conceptual background of self-organization by reactive processes. *Biol. Cell* **98**, 589–602 (2006).
- Kolmogorov, A., Petrovsky, L. & Piskunov, N. An investigation of the diffusion equation combined with an increase in mass and its application to a biological problem. *Bull. Uni. Moscow Ser. Int. A1* **6**, 1–26 (1937).
- Turing, A. M. The chemical basis of morphogenesis. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **237**, 37–72 (1952).
- Prigogine, I. & Nicolis, G. On symmetry-breaking instabilities in dissipative systems. *J. Chem. Phys.* **46**, 3542–3550 (1967).
- Prigogine, I., Nicolis, G. & Babloyantz, A. Nonequilibrium problems in biological phenomena. *Ann. NY Acad. Sci.* **231**, 99–105 (1974).
- Nicolis, G. & Prigogine, I. *Self-Organization in Nonequilibrium Systems: From Dissipative Structures to Order Through Fluctuations* (Wiley, New York, 1977).
- Goldbeter, A. & Lefever, R. Dissipative structures for an allosteric model. Application to glycolytic oscillations. *Biophys. J.* **12**, 1302–1315 (1972).
- Boiteux, A., Hess, B. & Plesser, T. Oscillatory phenomena in biological systems. *FEBS Lett.* **75**, 1–4 (1977).
- Goldbeter, A. *Biochemical Oscillations and Cellular Rhythms: The Molecular Bases of Periodic and Chaotic Behaviour* (Cambridge Univ. Press, 1996).
- Maini, P. K., Baker, R. E. & Chuong, C. M. Developmental biology. The Turing model comes of molecular age. *Science* **314**, 1397–1398 (2006).
- Murray, J. (ed.) *Mathematical Biology* (Springer, New York, 2007).
- Kauffman, S. *At Home in the Universe* (Oxford Univ. Press, 1995).
- Goodwin, B. C., Kauffman, S. & Murray, J. D. Is morphogenesis an intrinsically robust process? *J. Theor. Biol.* **163**, 135–144 (1993).
- Kauffman, S. *The Origins of Order: Self-Organization and Selection in Evolution* (Oxford Univ. Press, 1993).
- Ball, P. *The Self-Made Tapestry* (Oxford Univ. Press, 1999).
- Camazine, S. et al. *Self-Organization in Biological Systems* (Princeton Univ. Press, 2001).
- Karsenti, E., Newport, J., Hubble, R. & Kirschner, M. Interconversion of metaphase and interphase microtubule arrays, as studied by the injection of centrosomes and nuclei into *Xenopus* eggs. *J. Cell Biol.* **98**, 1730–1745 (1984).
- Kirschner, M. & Mitchison, T. Beyond self-assembly: from microtubules to morphogenesis. *Cell* **45**, 329–342 (1986).
- Tabony, J. & Job, D. Spatial structures in microtubular systems requiring a sustained energy source. *Nature* **346**, 448–451 (1990).
- Verde, F., Berrez, J. M., Antony, C. & Karsenti, E. Taxol induced microtubule asters in mitotic extracts of *Xenopus* eggs: requirement for phosphorylated factors and cytoplasmic dynein. *J. Cell Biol.* **112**, 1177–1187 (1991).
- Karsenti, E. Mitotic spindle morphogenesis in animal cells. *Semin. Cell Biol.* **2**, 251–260 (1991).
- Mitchison, T. J. Self-organization of polymer-motor systems in the cytoskeleton. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **336**, 99–106 (1992).
- Misteli, T. The concept of self-organization in cellular architecture. *J. Cell Biol.* **155**, 181–185 (2001).
- Kruse, K. & Jülicher, F. Oscillations in cell biology. *Curr. Opin. Cell Biol.* **17**, 20–26 (2005).
- Glick, B. S. Let there be order. *Nature Cell Biol.* **9**, 130–132 (2007).
- Bénard, H. Les tourbillons cellulaires dans une nappe liquide. *Rev. Gen. Sci. Pure Appl.* **11**, 1261–1271 (1900) (in French).
- Rayleigh, L. On convective currents in a horizontal layer of fluid when the higher temperature is on the under side. *Philos. Mag.* **32** (1916).
- Castets, V. V., Dulos, E., Boissonade, J. & De Kepper, P. Experimental evidence of a sustained standing Turing-type nonequilibrium chemical pattern. *Phys. Rev. Lett.* **64**, 2953–2956 (1990).
- Ouyang, Q. & Swinney, H. Transition from a uniform state to hexagonal and striped Turing patterns. *Nature* **352**, 610–612 (1991).
- Shoji, H., Yamada, K., Ueyama, D. & Ohta, T. Turing patterns in three dimensions. *Phys. Rev. E Stat. Nonlin. Soft Matter Phys.* **75**, 046212 (2007).
- Yang, L. & Epstein, I. Oscillatory Turing patterns in reaction-diffusion systems with two coupled layers. *Physic. Rev. Lett.* **90**, 178303 (2003).
- Gierer, A. & Meinhardt, H. A theory of biological pattern formation. *Kybernetik* **12**, 30–39 (1972).
- Meinhardt, H. & Gierer, A. Pattern formation by local self-activation and lateral inhibition. *Bioessays* **22**, 753–760 (2000).
- Tyson, J. J., Chen, K. C. & Novak, B. Sniffers, buzzers, toggles and blinkers: dynamics of regulatory and signaling pathways in the cell. *Curr. Opin. Cell Biol.* **15**, 221–231 (2003).
- Glade, N., Demongeot, J. & Tabony, J. Comparison of reaction-diffusion simulations with experiment in self-organized microtubule solutions. *CR Biol.* **325**, 283–294 (2002).
- Guo, Y., Liu, Y., Tang, J. X. & Valles, J. M. Polymerization force driven buckling of microtubule bundles determines the wavelength of patterns formed in tubulin solutions. *Phys. Rev. Lett.* **98**, 198103-1-4 (2007).
- Cortes, S., Glade, N., Chartier, I. & Tabony, J. Microtubule self-organisation by reaction-diffusion processes in miniature cell-sized containers and phospholipid vesicles. *Biophys. Chem.* **120**, 168–177 (2006).
- Maly, I. V. & Borisov, G. G. Self-organization of treadmilling microtubules into a polar array. *Trends Cell Biol.* **12**, 462–465 (2002).
- Nédélec, F. J., Surrey, T., Maggs, A. C. & Leibler, S. Self-organization of microtubules and motors. *Nature* **389**, 305–308 (1997).
- Surrey, T., Nédélec, F., Leibler, S. & Karsenti, E. Physical properties determining self-organization of motors and microtubules. *Science* **292**, 1167–1171 (2001).
- Nédélec, F. Computer simulations reveal motor properties generating stable antiparallel microtubule interactions. *J. Cell Biol.* **158**, 1005–1015 (2002).
- Nogales, E., Whittaker, M., Milligan, R. A. & Downing, K. H. High-resolution model of the microtubule. *Cell* **96**, 79–88 (1999).
- Vallee, R. B. & Stehman, S. A. How dynein helps the cell find its center: a servomechanical model. *Trends Cell Biol.* **15**, 288–294 (2005).
- Backouche, F., Haviv, L., Grosswasser, D. & Bernheim-Groszouche, A. Active gels: dynamics of patterning and self-organization. *Phys. Biol.* **3**, 264–273 (2006).
- Kruse, K., Joanny, J., Jülicher, F., Prost, J. & Sekimoto, K. Asters, vortices, and rotating spirals in active gels of polar filaments. *Phys. Rev. Lett.* **92**, 078101–078104 (2004).
- Haviv, L. et al. Reconstitution of the transition from lamellipodium to filopodium in a membrane-free system. *Proc. Natl Acad. Sci. USA* **103**, 4906–4911 (2006).
- Jülicher, F. & Prost, J. Spontaneous oscillations of collective molecular motors. *Phys. Rev. Lett.* **78**, 4510–4513 (1997).
- Goldbeter, A. Oscillations and waves of cyclic AMP in *Dictyostelium*: a prototype for spatio-temporal organization and pulsatile intercellular communication. *Bull. Math. Biol.* **68**, 1095–1109 (2006).
- Novak, B. & Tyson, J. J. Modelling the controls of the eukaryotic cell cycle. *Biochem. Soc. Trans.* **31**, 1526–1529 (2003).
- Sha, W. et al. Hysteresis drives cell-cycle transitions in *Xenopus laevis* egg extracts. *Proc. Natl Acad. Sci. USA* **100**, 975–980 (2003).
- Murray, A. W., Solomon, M. J. & Kirschner, M. W. The role of cyclin synthesis and degradation in the control of maturation promoting factor activity. *Nature* **339**, 280–286 (1989).
- Murray, A. W. & Kirschner, M. W. Dominoes and clocks: the union of two views of cell cycle regulation. *Science* **246**, 614–621 (1989).
- Félix, M.-A., Pines, J., Hunt, T. & Karsenti, E. Temporal regulation of Cdc2 mitotic kinase activity and cyclin degradation in cell-free extracts of *Xenopus* eggs. *J. Cell Sci.* **12**, 99–116 (1989).
- Félix, M. A., Labbé, J. C., Dorée, M., Hunt, T. & Karsenti, E. Triggering of cyclin degradation in interphase extracts of amphibian eggs by Cdc2 kinase. *Nature* **346**, 379–382 (1990).

73. Yang, L., MacLellan, W. R., Han, Z., Weiss, J. N. & Qu, Z. Multisite phosphorylation and network dynamics of cyclin-dependent kinase signaling in the eukaryotic cell cycle. *Biophys. J.* **86**, 3432–3443 (2004).
74. Zwolak, J. W., Tyson, J. J. & Watson, L. T. Parameter estimation for a mathematical model of the cell cycle in frog eggs. *J. Comput. Biol.* **12**, 48–63 (2005).
75. Hetzer, M., Gruss, O. J. & Mattaj, I. W. The Ran GTPase as a marker of chromosome position in spindle formation and nuclear envelope assembly. *Nature Cell Biol.* **4**, E177–E184 (2002).
76. Karsenti, E. & Vernos, I. The mitotic spindle: a self-made machine. *Science* **294**, 543–547 (2001).
77. Cook, P. R. Predicting three-dimensional genome structure from transcriptional activity. *Nature Genet.* **32**, 347–352 (2002).
78. Iborra, F. J. & Cook, P. R. The interdependence of nuclear structure and function. *Curr. Opin. Cell Biol.* **14**, 780–785 (2002).
79. Meaburn, K. J., Misteli, T. & Soutoglou, E. Spatial genome organization in the formation of chromosomal translocations. *Semin. Cancer Biol.* **17**, 80–90 (2007).
80. Misteli, T. Beyond the sequence: cellular organization of genome function. *Cell* **128**, 787–800 (2007).
81. Piekny, A., Werner, M. & Glotzer, M. Cytokinesis: welcome to the Rho zone. *Trends Cell Biol.* **15**, 651–658 (2005).
82. Camalet, S. & Jülicher, F. Generic aspects of axonemal beating. *New J. Phys.* **2**, 24.1–24.23 (2000).
83. Verde, F., Mata, J. & Nurse, P. Fission yeast cell morphogenesis: identification of new genes and analysis of their role during the cell cycle. *J. Cell Biol.* **131**, 1529–1538 (1995).
84. Brunner, D. & Nurse, P. New concepts in fission yeast morphogenesis. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **355**, 873–877 (2000).
85. Carazo-Salas, R. E. & Nurse, P. Self-organization of interphase microtubule arrays in fission yeast. *Nature Cell Biol.* **8**, 1102–1107 (2006).
86. Daga, R. R., Lee, K. G., Bratman, S., Salas-Pino, S. & Chang, F. Self-organization of microtubule bundles in nucleate fission yeast cells. *Nature Cell Biol.* **8**, 1108–1113 (2006).
87. Janson, M. E. *et al.* Crosslinkers and motors organize dynamic microtubules to form stable bipolar arrays in fission yeast. *Cell* **128**, 357–368 (2007).
88. Carazo-Salas, R. & Nurse, P. Sorting out interphase microtubules. *Mol. Syst. Biol.* **3**, 95 (2007).
89. Castagnetti, S., Novak, B. & Nurse, P. Microtubules offset growth site from the cell centre in fission yeast. *J. Cell Sci.* **120**, 2205–2213 (2007).
90. Devreotes, P. N. & Zigmond, S. H. Chemotaxis in eukaryotic cells: a focus on leukocytes and *Dictyostelium*. *Annu. Rev. Cell Biol.* **4**, 649–686 (1988).
91. Devreotes, P. & Janetopoulos, C. Eukaryotic chemotaxis: distinctions between directional sensing and polarization. *J. Biol. Chem.* **278**, 20445–20448 (2003).
92. Wedlich-Soldner, R., Wai, S. C., Schmidt, T. & Li, R. Robust cell polarity is a dynamic state established by coupling transport and GTPase signaling. *J. Cell Biol.* **166**, 889–900 (2004).
93. Xu, J. *et al.* Divergent signals and cytoskeletal assemblies regulate self-organizing polarity in neutrophils. *Cell* **114**, 201–214 (2003).
94. Maly, I. V., Wiley, H. S. & Lauffenburger, D. A. Self-organization of polarized cell signaling via autocrine circuits: computational model analysis. *Biophys. J.* **86**, 10–22 (2004).
95. Wedlich-Soldner, R. & Li, R. Spontaneous cell polarization: undermining determinism. *Nature Cell Biol.* **5**, 267–270 (2003).
96. Nigg, E. A. Centrosome duplication: of rules and licenses. *Trends Cell Biol.* **17**, 215–221 (2007).
97. Dutcher, S. K. Finding treasures in frozen cells: new centriole intermediates. *Bioessays* **29**, 630–634 (2007).
98. Vldar, E. K. & Stearns, T. Molecular characterization of centriole assembly in ciliated epithelial cells. *J. Cell Biol.* **178**, 31–42 (2007).
99. Pelletier, L. Centrioles: duplicating precariously. *Curr. Biol.* **17**, R770–R773 (2007).
100. Bornens, M. & Karsenti, E. In *Membrane Structure and Function* (ed. Bittar, E. E.) 99–171 (Wiley, New York, 1984).
101. Rodrigues-Martins, A., Riparbelli, M., Callaini, G., Glover, D. M. & Bettencourt-Dias, M. Revisiting the role of the mother centriole in centriole biogenesis. *Science* **316**, 1046–1050 (2007).
102. Azimzadeh, J. & Bornens, M. Structure and duplication of the centrosome. *J. Cell Sci.* **120**, 2139–2142 (2007).
103. Heald, R. & Weis, K. Spindles get the ran around. *Trends Cell Biol.* **10**, 1–4 (2000).
104. Pelletier, L. *et al.* Golgi biogenesis in *Toxoplasma gondii*. *Nature* **418**, 548–552 (2002).
105. He, C. Y., Pypaert, M. & Warren, G. Golgi duplication in *Trypanosoma brucei* requires Centrin2. *Science* **310**, 1196–1198 (2005).
106. He, C. Y. *et al.* Golgi duplication in *Trypanosoma brucei*. *J. Cell Biol.* **165**, 313–321 (2004).
107. Thiele, C. & Huttner, W. B. Protein and lipid sorting from the trans-Golgi network to secretory granules — recent developments. *Semin. Cell Dev. Biol.* **9**, 511–516 (1998).
108. Glick, B. S. Can the Golgi form *de novo*? *Nature Rev. Mol. Cell Biol.* **3**, 615–619 (2002).
109. Simpson, J. C., Nilsson, T. & Pepperkok, R. Biogenesis of tubular ER-to-Golgi transport intermediates. *Mol. Biol. Cell* **17**, 723–737 (2006).

Acknowledgements

I thank colleagues of the Cell Biology and Biophysics Unit at the European Molecular Biology Laboratory for all of the discussions and collective work. I also thank the three anonymous reviewers who helped bring this text to something that looks, I hope, like a balanced and coherent whole. I apologize to all those who have not been cited owing to lack of space or because of my ignorance.

DATABASES

UniProtKB: <http://beta.uniprot.org/uniprot>
CDK1 | Orb6 | PLK4 | Ran | RhoA

FURTHER INFORMATION

Eric Karsenti's homepage:

http://www-db.embl.de/jss/EmblGroupsOrg/g_40.html

Rayleigh number: <http://scienceworld.wolfram.com/physics/RayleighNumber.html>

ALL LINKS ARE ACTIVE IN THE ONLINE PDF